



EMERGING INFECTIOUS DISEASES

Current Issue

Vol. 8, No. 1
January 2002

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Letter

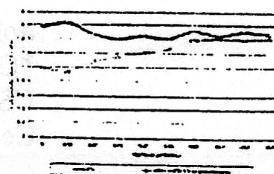
Efficacy of Interferon α -2b and Ribavirin against West Nile Virus In Vitro

To the Editor: *West Nile virus* (WNV) infected humans in the Western Hemisphere for the first time in the late summer of 1999. During 1999 and 2000, nine deaths occurred among 80 patients with meningitis or encephalitis in New York City; Westchester County, New York; New Jersey; and Connecticut (1-3). Effective antiviral agents are unknown for infections caused by WNV. Odelola (4) described 83% survival of WNV-infected mice and eradication of virus from brain when 1.5 mg of ribavirin was administered by intraperitoneal injection after virus inoculation. Survival of controls was 25%. More recently, Jordan et al. have reported inhibition of WNV by a relatively high concentration of ribavirin (200 M) given after infection of human oligodendroglial cells in vitro (5). Shahar et al. (6) reported protection of fetal mouse spinal cord tissues with mouse alpha and beta interferon before inoculation with WNV. We tested human recombinant interferon alpha-2b and ribavirin for activity against WNV in a primate cell system similar to that used to measure the effect of these agents on *Bovine viral diarrhea virus*, a cultivatable, closely related surrogate for *Hepatitis C virus*.

Vero cells were cultured at 37° and 5% CO₂ in a 96-well microtiter plate. Approximately 13,000 cells were seeded in each well 24 hours before specific concentrations of ribavirin or interferon alpha-2b were added. Approximately 2.5 X 10³ PFU of WNV isolated from *Culex pipiens* (7) was added 1.5-2 hours after or before the addition of interferon alpha-2b or ribavirin to Vero cells. Forty-four hours after treatment, a colorimetric proliferation assay was used to measure viable cells in each treated well according to the protocol of Promega (Madison, WI). Cells exposed to specific concentrations of antiviral compounds, but without WNV, were used as negative controls.

Interferon alpha-2b was protective and therapeutic. Interferon alpha-2b inhibited viral cytotoxicity at low dosage when applied before or after infection of cells with WNV. For example, viral protection was observed from 3,000 U/mL to 188 U/mL 2 hours before infection of cells with WNV. Interferon alpha-2b

Figure



was also therapeutic when applied after cells were infected with WNV. Virus-induced cytotoxicity was inhibited by concentrations of ≥ 5.9 U/mL when added 1.5 hours after infection (Figure). The optical density 490 values in these tests were significantly different ($p < 0.05$, using Tukey HSD multiple comparison test) when compared with the uninfected cells.

Ribavirin was protective but not therapeutic in vitro. Cells were protected at dosages of 400 and 500 μ M but not at dosages of 300 μ M of ribavirin applied 2 hours before infection of cells with WNV. A cytotoxic effect of ribavirin occurred at concentrations of 600-1,000 μ M.

In humans, daily doses of 3 million units of interferon result in serum levels of 10-20 U/mL, well above that required for in vitro efficacy (8). In contrast, oral ribavirin doses of 2,400 mg daily yield a steady-state serum concentration of 3-4 μ g/mL after several days, approximately 12-40-fold less than the in vitro inhibitory concentrations of 200-500 M (50-125 μ g/mL) noted by Jordan et al. (5) and in this study. Intravenous administration of 4 g daily, as used in the treatment of Lassa fever, would be required to reach a potentially effective serum concentration (9,10). However, intracellular accumulation and phosphorylation of ribavirin may account for its therapeutic effect in mice (4).

We conclude that interferon alpha-2b possesses greater therapeutic activity in vitro than ribavirin, with a potentially greater therapeutic ratio in humans. Whether combination therapy, as employed against hepatitis C, may be optimal requires further study.

Acknowledgments

Schering-Plough Research Institute provided technical quantities of interferon alpha-2b and ribavirin. Paul Glue, Paul Ingravallo, and Gerald Hajian provided helpful information. Jodi Correia, Bonnie Hamid, and Jeffrey Ward provided technical assistance. Noriel Mariano assisted in preparing the manuscript. Dr. Johnson Y.N. Lau collaborated in protocol development.

This study was supported in part by the BMA Medical Foundation, the Beatrice Snyder Foundation, the Hugaton Foundation, and the U.S. Department of Agriculture, Specific Cooperative Agreement 58-6615-1-20.

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Figure. Effect of varying concentrations of interferon alpha-2b (FN) on West Nile virus-infected Vero cells. The vertical axis represents a colorimetric assay of cellular lactic dehydrogenase, which is directly proportional to cell viability and proliferation. OD = optical density.



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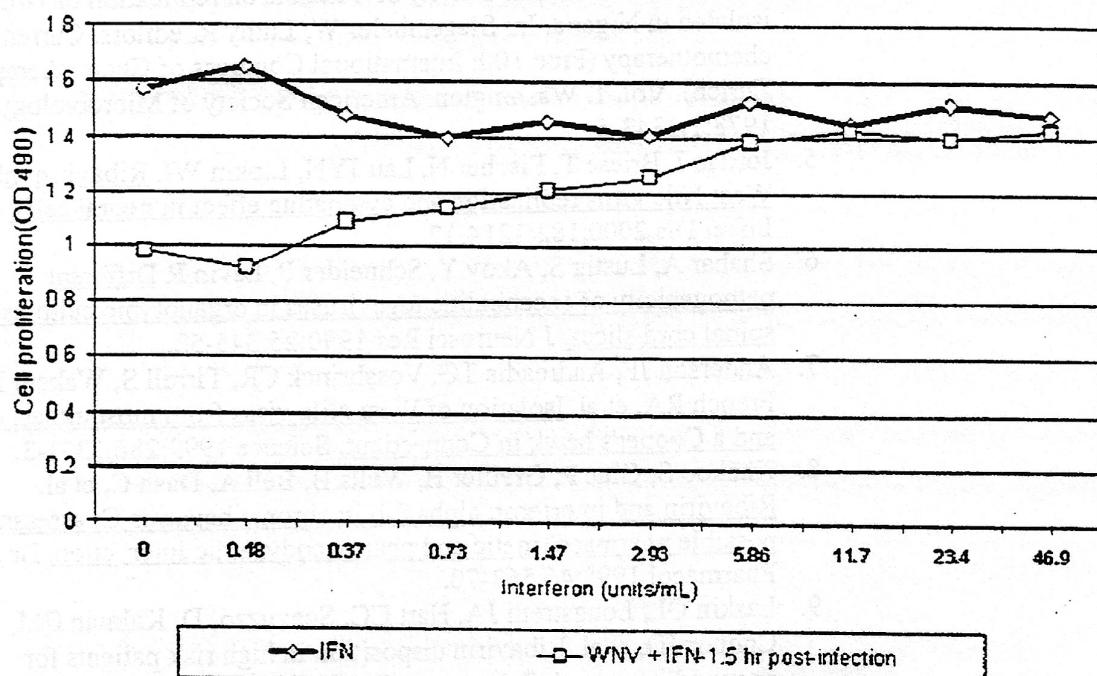
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This page last reviewed December 07, 2001

Emerging Infectious Diseases Journal
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References

1. Centers for Disease Control and Prevention. Guidelines for surveillance, prevention, and control of West Nile Virus infection—United States. MMWR Morb Mortal Wkly Rep 2000;49:25-8
2. Layton M. West Nile virus final update for the 2000 season. New York: City of New York Department of Health Communicable Disease Program; 2000. p. 1-4.
3. Lanciotti RS, Roehrig JT, Deubel V, Smith J, Parker M, Steele K, et al. Origin of the West Nile virus responsible for an outbreak of encephalitis in the Northeastern United States. Science 1999;286:2333-7.
4. Odelola HA. Antiviral activity of Virazole on replication of viruses isolated in Nigeria. In: Siegenthaler W, Luthy R, editors. Current chemotherapy (Proc 10th International Congress of Chemotherapy, Zurich). Vol. 1. Washington: American Society of Microbiology; 1978;1:3343-5.
5. Jordan I, Briese T, Fischer N, Lau JYN, Lipkin WI. Ribavirin inhibits West Nile virus replication and cytopathic effect in neural cells. J Infect Dis 2000;182:1214-17.
6. Shahar A, Lustig S, Akov Y, Schneider P, Levin R. Different pathogenicity of encephalitic togaviruses in organotypic cultures of spinal cord slices. J Neurosci Res 1990;25:345-52.
7. Anderson JF, Andreadis TG, Vossbrinck CR, Tirrell S, Wakem EM, French RA, et al. Isolation of West Nile virus from mosquitoes, crows, and a Cooper's hawk in Connecticut. Science 1999;286:2331-3.
8. Khakoo S, Glue P, Grellier H, Wells B, Bell A, Dash C, et al. Ribavirin and interferon alpha 2-b in chronic hepatitis C: assessment of possible pharmacokinetic and pharmacodynamic interaction. Br J Clin Pharmacol 1998;46:563-70.
9. Laskin OL, Lougheed JA, Hart CC, Scavuzzo, D, Kalman CM, Connor JD, et al. Ribavirin disposition in high risk patients for acquired immunodeficiency syndrome. Clin Pharmacol Ther 1987;41:546-55.
10. McCormick JB, King IJ, Webb PA, Scribner CL, Craven RB, Johnson KM, et al. Lassa fever. Effective therapy with ribavirin. N Engl J Med 1986;314:20-6.

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